



World Scientific News

An International Scientific Journal

WSN 108 (2018) 180-194

EISSN 2392-2192

Effect of Extract-Fractions of *Balanites aegyptiaca* Fruit-Mesocarp on Glucose Metabolizing Enzymes in Diabetic Rats

Daniel Hassan Mhya^{1,*}, Janet O. Alegbejo², Kola Matthew Anigo³,
Ismail Alhaji Umar³

¹Department of Biochemistry, Abubakar Tafawa Balewa University Bauchi, PMB 0248, Bauchi, Nigeria

²Department of Pediatric, Ahmadu Bello University Teaching Hospital Zaria, PMB 06 Shika, Nigeria

³Department of Biochemistry, Ahmadu Bello University Zaria, PMB 1045 Nigeria

*E-mail address: dmhassan@atbu.edu.ng

*Phone: +2348039652964

ABSTRACT

Several antidiabetic medicinal plants has shown to exhibit one of more mechanisms to low blood glucose. *Balanites aegyptiaca* fruit extract has shown to low blood glucose via some of the mechanisms but whether the fruit extract could stimulate glycolysis, glycogenesis, and inhibit gluconeogenesis and glycogenolysis are not fully clear. Hence, the need to evaluate the plant extract on the activities of enzymes in glycolysis, gluconeogenesis, glycogenolysis and glycogenesis in streptozotocin-induced diabetic rats. Ethanol extract of defatted *Balanites aegyptiaca* fruit-mesocarp was petition with water and ethyl acetate (1:1 v/v) then separated. Aqueous and ethyl acetate fractions obtained were separately administered to streptozotocin-induced diabetic rats daily for 28 days period. Results of the study showed that treating diabetic rats with either aqueous or ethyl acetate fractions of ethanol extract of *Balanites aegyptiaca* fruit-mesocarp exert significant regulatory effect on the activities of some key hepatic enzymes of glucose metabolism. The aqueous fraction (AFF) in particular enhances glucokinase activity (from 2.22 ± 0.02 to 3.58 ± 0.05 U/min/mg protein) and G6PDH activity (from 1.45 ± 0.02 to 2.10 ± 0.02 U/min/mg protein) but suppressed glucose-6-phosphatase (from 1.44 ± 0.05 to 0.17 ± 0.00 U/min/ μ mole Pi liberated) among others. In conclusion, ethanol-aqueous fraction of *Balanites aegyptiaca* fruit-mesocarp exert glucose enzymes regulatory effect by enhancing glucokinase and G6PDH activity but suppressed gluconeogenic enzymes possibly to low blood

glucose in diabetic rats. Further research is needed to explore the plant extract (ethanol-aqueous fraction) in order to ascertain the bioactive hidden compounds.

Keywords: *Balanites aegyptiaca*, Fruit-Mesocarp, Ethanol Extract-Fractions, glucose metabolizing enzymes

1. INTRODUCTION

Hyperglycemia in diabetes mellitus is largely results from hepatic glucose over production associated with glucose metabolic disorder which occurs as a result of disturbance in the activities of enzymes involved in glycolysis, gluconeogenesis, glycogenesis and glycogenolysis [1]. Studies have shown that disturbance of carbohydrate metabolism has significant influences on glucose homeostasis [2,3]. Deficiency of insulin disrupts carbohydrate metabolism by suppressing the activities of glycolytic and glycogenic enzymes while promoting gluconeogenic and glycogenolytic enzymes [1,4].

Maintenance of normal glycemia requires matching of glucose utilization and endogenous production. This could be achieve via coordinated regulations of several metabolic pathways; glycolysis, gluconeogenesis, glycogenolysis, and glycogenesis [5,6]. Several regulatory enzymes like glucokinase, phosphofructokinase, pyruvate kinase, glucose-6-phosphatase, fructose-1,6-bisphosphatase, phosphoenolpruvate carboxlkinase, glycogen synthase and glycogen phosphorylase play key roles in these metabolic pathways [6].

According to Abdollahi *et al* [7], any agent with potential to reverse hepatic carbohydrate metabolism might have influence on enzymes involved in glucose and glycogen metabolisms. To this regard, Agius [8] has reported the inhibition of enzymes involved in gluconeogenesis and glycogenolysis which led to the lowering of fasting plasma glucose. Some antidiabetic medicinal plants has shown significant impacts by modulating the activities of glucose metabolizing enzymes [9,10].

Studies has reported that plants extract reverses phosphoenolpyruvate carboxykinase (PEPCK) and glucose 6-phosphatase activities [11,12].

The plant '*Balanites aegyptiaca Delile*', also known as 'desert date' in English, a member of *Zygophyllaceae* family, is a common plant specie of the dry land areas of Africa and Asia [13,14]. In Nigeria, it is found mostly in the Northern region. It is known as '*Aduwa*' in Hausa, '*Utazi*' in Igbo, and '*Teji*' in Yoruba. *Balanites aegyptiaca* has a long history of traditional uses for wide ranges of disease [15].

The fruit extracts of *Balanites aegyptiaca* was reported to have exhibited prominent antihyperglycemic activity in diabetic-induced animals [16-18]. It was reported that the plant fruit extract stimulated insulin secretion [16,19], increased muscle basal glucose uptake [20] to lowered blood glucose level. It was also reported that it retards the activities of some enzymes of carbohydrate metabolism such as intestinal α -amylase [21]. In a recent studies, the plant leaves extract was reported to have inhibited alpha amylase activity *in vitro* [22] while in our study it was found to reverse the activities of some key hepatic enzymes of glucose metabolism in diabetic rats [23].

It was reported that the antidiabetic activity of medicinal plants depends upon a variety of mechanisms which include: stimulation of insulin secretion, inhibition of insulin degradative processes and reduction of insulin resistance [24], regenerating or repairing

pancreatic β -cells and protecting the destruction of the β -cells [25], stimulation of glycogenesis and hepatic glycolysis [26], inhibition of α -amylase/ α -glucosidase enzymes [27], inhibition of gluconeogenesis [8] and preventing oxidative stress in pancreatic β - cell dysfunction [28]. *Balanites aegyptiaca* fruit extract has shown to low blood glucose via some of the mechanisms but whether the fruit extract could stimulate glycolysis, glycogenesis, and inhibit gluconeogenesis and glycogenolysis are not fully clear. Hence, the need to evaluate the plant extract on the activities of enzymes in glycolysis, gluconeogenesis, glycogenolysis and glycogenesis in streptozotocin-induced diabetic rats.

2. MATERIALS AND METHODS

2. 1. Materials

a) Chemicals/reagents

All chemicals/reagents used were of analytical grade and were obtained from Sigma Aldrich, USA; British Drug House, England; Randox laboratory, UK.

b) Plant collection

Balanites aegyptiaca fruit-mesocarp was collected from Gubi village (latitude 10° 45' N & longitude 9° 82' E) in Bauchi, Bauchi state. It was identified and authenticated at the Herbarium Unit, Department of Biological Science, Ahmadu Bello University Zaria. A voucher specimen (voucher no: 900175) was deposited in the herbarium of the Department.

2. 2. Experimental animals

A total of twenty-five (25) male wistar albino rats were used for the study. The rats were obtained from the Animal House, University of Jos, Plateau State, Nigeria and kept in clean cages with 12 hours / 12 hours light/dark photoperiod. Water and feed 'growers mash' (Vital feeds, Jos) were supplied *ad libitum*. The rats were allow to grow attaining a weight between 180-230g before used. All experimental protocol was in conformity with the institutional guidelines that are in compliance with national and international laws and guidelines for care and use of laboratory animals [29].

2. 3. Methods

a) Plant extraction/fractionation

Plant fruit-mesocarp was defatted as performed by Jung *et al* [30] and extracted as done by Govorko *et al* [31] with little modification in the choice of the extraction temperature (60 °C). Seven hundred and fifty gram (750g) powdered of plant fruit-mesocarp was defatted for 2 hours with 1200 ml hexane on a mechanical shaker.

The hexane solvent was discarded, then the defatted sample air-dried. Exactly 200 g of the defatted plant fruit-mesocarp was mixed with 2000 ml of 80 % ethanol and heated to 60 °C for 2 hours. The extraction continued for an additional 10 hrs at 20 °C. The mixture was filtered through a cheese cloth and resulting ethanol extract was air-dried. The procedure was repeated twice with same amount of defatted plant fruit-mesocarp.

Ethanol fruit-mesocarp extract was dissolved in water (500 ml) and partitioned with ethyl acetate (500 ml) at 20 °C for 2 hours then separated using a separating funnel (1000 ml). Fractions were concentrated using a rotary evaporator at 40 °C and air dried. The dried aqueous (AFF) and ethyl acetate (EFF) fractions of *Balanites aegyptiaca* fruit-mesocarp were stored in air-tight containers and kept in a refrigerator at 4 °C until used.

b) Induction of diabetes mellitus

Diabetes mellitus was induced in rats by intra-peritoneal injection of Streptozotocin (STZ) at a dose of 60 mg/kg body wt dissolved in 0.1 M citrate buffer (pH 4.5). Rats were given 10 % glucose solution in their drinking water for 48 hours after STZ injection in order to prevent severe hypoglycemia. After 72 hours, blood glucose levels were checked and subsequent 1-week intervals to identify the onset and continued presence of diabetic hyperglycemia; rats with fasting blood glucose levels ≥ 200 mg/dl were considered diabetic and selected for the study [32].

2. 4. Experimental design

Effects of ethanolic extract-fractions of *Balanites aegyptiaca* fruit-mesocarp on the activities of glucose metabolic enzymes was assessed in the streptozotocin-induced diabetic rats. Rats were randomly allocated into groups of 5 rats each as follows;

- Group A : Diabetic + Aqueous fruit-Mesocarp fraction (AFF)
- Group B : Diabetic + Ethyl acetate fruit-Mesocarp fraction (EFF)
- Group C : Diabetic + Metformin at 200 mg/kg body weight (kolawole and Akanji [12].
- Group E : Diabetic control
- Group F : Normal control

At the end of the experiment, animals were sacrificed, liver were excised, homogenized and was used for the assay of the glucose metabolic enzymes. The extract-fractions were administered orally using oral gastric tube. Exact 400 mg/kg body weight of plant extracts were administered to various diabetic rats' groups daily for 28 days period. The extract-dose used was determined following our previous acute toxicity report on the AFF and EFF of *Balanites aegyptiaca* [33].

2. 5. Assay of glucose metabolic enzymes' activity

Hepatic key glucose metabolic enzymes like Glucokinase [34], Phosphofructokinase [35], fructose-1,6-bisphosphatase [36], Phosphoenolpyruvate carboxylkinase [37], Glucose-6-phosphate dehydrogenase [38], Glycogen phosphorylase activity [39], Glucose-6-phosphatase activity [40], Glycogen synthase activity [41], Pyruvate kinase [42] were all assayed following standard procedures.

2. 6. Statistical analysis

Data from the experiments were expressed as mean \pm standard deviation (SD). Means were analyzed by one way analysis of variance (ANOVA) and compared by Duncan's multiple range test (DMRT) [43]. Significant difference was accepted at $P < 0.05$.

3. RESULTS

3. 1. Effect of EFF and AFF on glycolytic enzymes in diabetic rats

Change in the activities of glucokinase, phosphofructokinase and pyruvate kinase assayed from liver of non-diabetic, diabetic treated and diabetic untreated rats is presented in Table 1. *Balanites aegyptiaca* fruit-mesocarp extract-fractions showed a stimulatory effect on the glycolytic enzymes activities particularly the aqueous-fraction (AFF) which significantly ($P < 0.05$) enhances glucokinase (from 2.22 ± 0.02 to 3.58 ± 0.05 U/min/mg protein).

Table 1. Effect of Ethanol Extract-Fractions of *Balanites aegyptiaca* Fruit-Mesocarp on Glycolytic Enzymes Activities in Liver of Streptozotocin-induced Diabetic Rats

| | Animal Grouping | | | | |
|--|------------------|-------------------|---------------------|-------------------------|-------------------|
| | Diabetic+ AFF | Diabetic + EFF | Diabetic Control | Diabetic + Metformin | Normal Control |
| Glucokinase (U/min/mg Protein) | 3.58± 0.05 | 3.10± 0.02 | 2.22± 0.02 | 2.72± 0.02 | 3.53± 0.01 |
| Phosphofructokinase (U/min/mg Protein) | 2.30± 0.57 | 1.25± 0.03 | 2.06± 0.07 | 3.34± 0.01 | 4.43± 0.08 |
| Pyruvate Kinase (U/min/mg Protein) $\times 10^{-1}$ | 0.30± 0.03 | 0.09± 0.03 | 0.04± 0.01 | 0.15± 0.02 | 0.11± 0.01 |
| LDH (U/min/mg Protein) $\times 10^{-2}$ | 6.07± 2.05 | 7.13± 2.69 | 4.36± 1.58 | 6.12± 2.19 | 7.10± 1.60 |

Values are Mean \pm SD of 5 determinations. Values with different superscript across the rows are significantly different ($P < 0.05$)

AFF = Aqueous Fruit-mesocarp fraction, EFF = Ethyl acetate fruit-mesocarp fraction, LDH = Lactate Dehydrogenase

3. 2. Effect of EFF and AFF on gluconeogenic enzymes in diabetic rats

Changes in the activities of hepatic gluconeogenic enzymes namely glucose-6-phosphatase, fructose-1,6-bisphosphatase, and phosphoenol pyruvate carboxyl kinase in diabetic untreated, diabetic treated and non-diabetic rats are shown in Table 2. The diabetic untreated rats showed an increase in gluconeogenic enzymes activity. However, these were significantly ($P < 0.05$) suppressed in the diabetic treated animals.

Aqueous fruit-mesocarp fraction (AFF) suppressed glucose-6-phosphatase (from 1.44±0.05 to 0.17±0.0 U/min/μmole P_i liberated) and phosphoenol pyruvate carboxylkinase (from 0.81±0.15 to 0.38±0.04 U/min/mg protein) while ethyl acetate fraction (EFF) suppressed fructose-1,6-bisphosphatase (from 2.19±0.25 to 1.20±0.03 U/min/μmole P_i liberated) among others.

Table 2. Effect of Ethanol Extract-Fractions of *Balanites aegyptiaca* Fruit-Mesocarp on Gluconeogenic Enzymes Activities in Liver of Streptozotocin-induced Diabetic Rats

| | Animal Grouping | | | | |
|---|------------------|-------------------|---------------------|-------------------------|-------------------|
| | Diabetic+ AFF | Diabetic + EFF | Diabetic Control | Diabetic + Metformin | Normal Control |
| Glucose-6-Phosphatase (U/min/μmole P _i liberated) | 0.17± 0.00 | 0.70± 0.01 | 1.44± 0.05 | 0.12± 0.02 | 0.07± 0.01 |
| Fructose-1,6-Bis-Phosphatase (U/min/μmole P _i liberated) | 1.41± 0.06 | 1.20± 0.03 | 2.19± 0.25 | 1.02± 0.02 | 1.40± 0.07 |
| Phosphoenol-pyruvate Carboxyl kinase (U/min/mg Protein) | 0.38± 0.04 | 0.46± 0.01 | 0.81± 0.15 | 0.11± 0.04 | 0.09± 0.01 |

Values are Mean ± SD of 5 determinations. Values with different superscript across the rows are significantly different (P<0.05)

AFF = Aqueous Fruit-mesocarp fraction, EFF = Ethyl acetate fruit-mesocarp fraction

3. 3. Effect of EFF and AFF on glycogenolytic and glycogenesis enzymes in diabetic rats

Activities of glycogen metabolic enzymes; glycogen synthase (GS) and phosphorylase in liver of diabetic untreated, non-diabetic and diabetic rats treated with ethyl acetate and aqueous fractions of *Balanites aegyptiaca* fruit-mesocarp are presented in Table 3.

A significant (P<0.05) decreased in glycogen synthase activity was recorded in the diabetic control rats group (9.41±0.34 × 10² U/min/mg protein) compared with metformin treated rats group (15.51±0.42 × 10² U/min/mg protein) and the plant ethanol extract-fractions; aqueous fraction (12.24±0.22 × 10² U/min/mg protein) and ethyl acetate fraction (11.75±0.11 × 10² U/min/mg protein). However, there was no significant difference (P>0.05) in the activity of glycogen phosphorylase from the diabetic treated and untreated diabetic rats.

Table 3. Effect of Ethanol Extract-Fractions of *Balanites aegyptiaca* Fruit-Mesocarp on Glycogen Content and Glycogen Metabolic Enzymes Activities in Liver of Streptozotocin-induced Diabetic Rats

| | Animal Grouping | | | | |
|---|------------------|-------------------|-----------------------------|----------------------------|-------------------|
| | Diabetic+ AFF | Diabetic + EFF | Diabetic Control | Diabetic + Metformin | Normal Control |
| Hepatic Glycogen (mg/g liver) | 20.62± 0.44 | 14.54± 0.32 | 10.69± 0.32 | 17.77± 0.32 | 15.85± 0.32 |
| Glycogen Phosphorylase (U/min/mg Protein) | 3.75± 0.00 | 3.46± 0.01 | 3.82± 0.21 ^{bc} | 2.04± 0.01 ^a | 2.07± 0.01 |
| Glycogen Synthase (U/min/mg Protein) ×10 ⁻² | 12.24± 0.22 | 11.75± 0.11 | 9.41± 0.34 | 15.51± 0.42 | 29.25± 0.88 |

Values are Mean ± SD of 5 determinations. Values with different superscript across the rows are significantly different (P<0.05)

AFF = Aqueous Fruit-mesocarp fraction, EFF = Ethyl acetate fruit-mesocarp fraction

4. 4. Effect of EFF and AFF on enzyme of hexomonophosphate pathway in diabetic rats

Activity of glucose-6-phosphate dehydrogenase (G6PDH) was determined in the liver tissues of diabetic treated rats in order to assess the impact of ethyl acetate and aqueous fractions of *Balanites aegyptiaca* fruit-mesocarp (Figure II). Treatment with metformin and ethanol extract-fractions of *Balanites aegyptiaca* fruit-mesocarp enhanced G6PDH activity by varying degrees. The aqueous fraction effectively enhanced G6PDH activity (from 1.45±0.02 to 2.10±0.02 U/min/mg protein) compared with values obtained from diabetic rats group treated with the ethyl acetate fraction.

5. DISCUSSION

Balanites aegyptiaca fruit extracts has been reported to exert potential antihyperglycemic activity [20,44]. Studies to explain how *Balanites aegyptiaca* fruit extract lowered fasting blood glucose suggested stimulation of insulin secretion [16], inhibition of intestinal α -amylase activity [21], increased muscle basal glucose uptake [16] as well as antioxidant activity [45]. Antidiabetic medicinal plants has been reported to exhibit variety of mechanisms to low blood glucose. The mode of actions exhibited by *Balanites aegyptiaca*

fruit extract might be attributable to the variety of different biologically active chemicals in the fruit [16,46,47].

In a recent studies, the plant leaves extracts was reported to have inhibited alpha amylase activity *in vitro* [22] and also reverses the activities of some key hepatic enzymes of glucose metabolism in diabetic rats [23]. Shafik *et al* [48] has reported that seed kernel of *Balanites aegyptiaca* enhanced glucokinase activity in diabetic rats. Activities of glucokinase, phosphofructokinase and pyruvate kinase has been shown to be

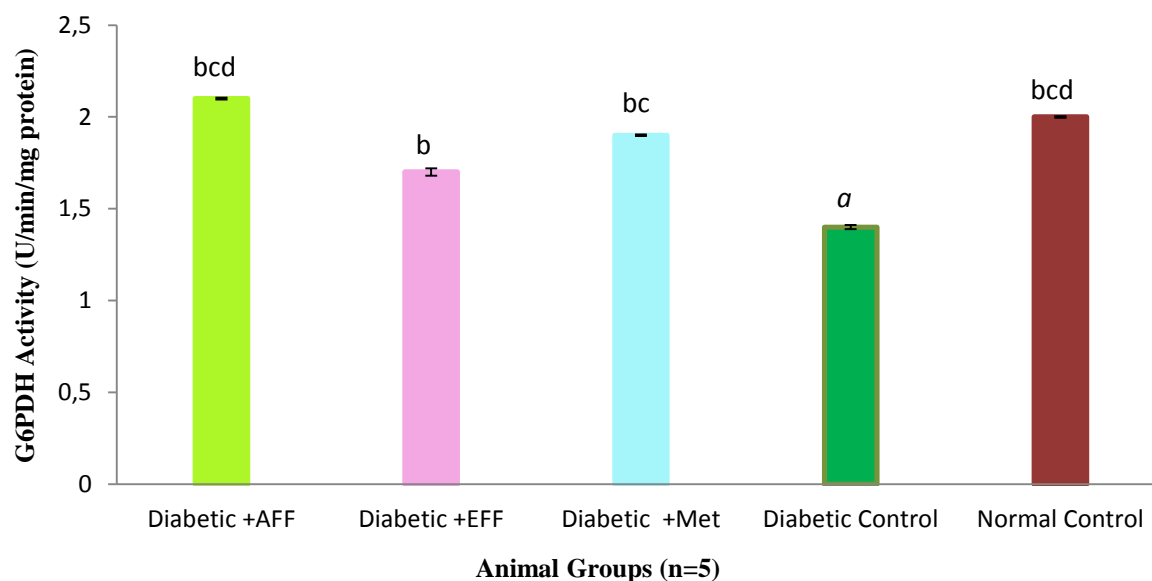


Figure I. Effect of Ethanol Extract-Fractions of *Balanites aegyptiaca* Fruit-Mesocarp on Glucose 6-phosphadehydrogenase Activity in STZ-induced Diabetic Rats

Bars with different letters are significantly different (P<0.05)

AFF = Aqueous Fruit-mesocarp fraction, EFF = Ethyl acetate fruit-mesocarp fraction very sensitive signs of the glycolytic pathway and these are decreased in the liver of diabetic state [49].

Reduced activities of these enzymes in this study are consistent with other studies on glycolytic enzymes [1,50]. Administration of fractions of the ethanolic extract of *Balanites aegyptiaca* fruit induced significant increase in the activities of glycolytic enzymes supporting the notion that part of the therapeutic potential of several putative antidiabetic plants can involve the modulation of enzymes in carbohydrate metabolism [51,52].

The activity of hepatic glucokinase is reported to be reduced in diabetes mellitus and can be activated by an activator [53]. Increased glucokinase activity in diabetic rats treated with extract-fractions of *Balanites aegyptiaca* fruit-mesocarp might have improve glycolysis leading to the reduction in blood glucose. Shafik *et al* [48] have shown that extract of *Balanites aegyptiaca* seed-kernel promotes the activity of hexokinase, suggesting glycolysis stimulatory effect of the plant.

Gluconeogenesis is a main cause of the elevated hepatic glucose production contributing 50-60 % of the released glucose [54]. It has been noted that metformin inhibit

hepatic gluconeogenesis to achieved anti-diabetic effect [55, 56]. *Balanites aegyptiaca* might have lowered blood glucose in part by inhibition of hepatic gluconeogenesis. Plants like *Eugenia jambolana* [57], *Centaurea bruguiera* ssp. *Belangerana* [58], and *Juglans regia* [59] have been reported to inhibit hepatic gluconeogenesis as part of their anti-diabetic mechanism.

In this study, extract-fractions of *Balanites aegyptiaca* fruit-mesocarp suppressed the activities of the gluconeogenic enzymes. This is in line with other studies where several plants extract were reported to have suppressed the activities of the gluconeogenic enzymes in diabetic animals [12,27,60,61]. Shafik *et al* [48] have reported that extract of *Balanites aegyptiaca* seed-kernel suppressed glucose-6-phosphatase activity.

Glycogen synthase (GS) catalyzes the rate limiting step in glycogen synthesis and is thus responsible for the storage of glucose as glycogen in the liver. Fractions of ethanolic extract of *Balanites aegyptiaca* fruit-mesocarp appear to has no effect on glycogen phosphorylase activity but activate synthase activity significantly in the diabetic treated rats. This is supported by the relative change in glycogen content in liver of the diabetic treated rats. Some plants extract have been reported to regulated glycogen enzymes leading to increased hepatic glycogen content [62,63]. According to Gutierrez [64], activation of glycogen synthase by plant suggested insulinogenic character; going by this statement one may propose that *Balanites aegyptiaca* contains component that exhibits insulin like effect.

Glucose-6-phosphate dehydrogenase is the first rate limiting enzyme of the pentose phosphate pathway which results in the production of ribose-5-phosphate and the reducing equivalent NADPH [65]. The study observed suppressed activity of G6PDH in liver of STZ-diabetic rats similar to reports by Karuna *et al* [66] and Saralakumari *et al* [67]. Diaz-Flores *et al* [68] have reported that decreased hepatic G6PDH activity depends on the severity of hyperglycemia. Decreased activity of G6PDH indicated low level of NADPH produced by hexose monophosphate pathway (HMP) which is unable to meet the cellular requirement for the enzymes that continuously maintain glutathione (GSH) in its reduced state or accumulation of glucose-6-phosphate which is a potent glycosylating agent that causes GSH depletion and thereby boosts glycation and it may also promote the final step of gluconeogenesis [69].

Balanites aegyptiaca fruit extract-fractions administration increased G6PDH activity in the diabetic treated rats, accompanied by increase in NADPH levels, a product of the HMP pathway. This might suggest that the ethanol-aqueous fraction of *Balanites aegyptiaca* fruit-mesocarp has the capacity to modulate hexose monophosphate pathway for alternative glucose oxidation. In addition, Atangwho *et al* [70] reported that glucose oxidation via gluconate pathway is associated with increased glucokinase activity. In this study, it is suggested that activation of G6PDH in the diabetic rats groups particularly that received the ethanol-aqueous fraction of *Balanites aegyptiaca* fruit-mesocarp may be a reflection of the increased activity of their glucokinase.

6. CONCLUSION

In conclusion, ethanol extract-fractions of *Balanites aegyptiaca* fruit-mesocarp could exert enzymes regulatory effect as recorded in this study from liver of diabetic rats. Aqueous fraction shows to be the most potent, it enhances glucokinase and glucose-6-

phosphatedehydrogenase (G6PDH) activity suggesting that the plant fruit extract could promote glucose oxidation to low blood glucose. Further research is needed to explore the fruit-mesocarp ethanol-aqueous fraction for its pharmaceutical importance.

ACKNOWLEDGEMENT

We acknowledge the efforts of Mallam Adamu Mohammed and Mr. Kabir Abdullahi both from the Department of Pharmacognosy, ABU Zaria, Nigeria for their technical assistance during the extraction/fractionations of plant sample.

References

- [1] Soliman MM, Ahmed MM, El-Shazly SM. Cinnamon extract regulates gene expression of lipid and carbohydrate metabolism in streptozotocin induced diabetic wistar rats. *American Journal of Biochemistry and Biotechnology* 9(2) (2013) 172-182.
- [2] Abdollahi, M., Donyavi, M., Pournourmohammadi, S., and Saadat, M. Hyperglycemia associated with increased hepatic glycogen phosphorylase and phosphoenolpyruvate carboxykinase in rats following subchronic exposure to Malathion. *Complimentary Biochemistry, Physiology and Toxicology Pharmacology*, 137(2004a) 343–347.
- [3] Atefi, M., Ghazanfari, S., Ostad, S.N., Ghahremani, M.H., and Abdollahi, M. Alteration of glucose homeostasis by rolipram and milrinone but not sildenafil in rat primary hepatocytes culture. *Progress in Medical Research*, 2(13) (2005) 1-12.
- [4] Skim, F., Lazrek, H.B., Kaaya, A., El-Amri, H., and Jana, M. Pharmacological studies of two antidiabetic plants: *Globulria alypun* and *Zypophyllum gaetulum*, *Therapie*, 54(6) (1999) 711-715.
- [5] Hers, H.G. Mechanisms of blood glucose homeostasis. *Journal of Inherit Metabolism Disease*, 13(4) (1999) 395-410.
- [6] Nordlie, R.C., Foster, J.D., and Lange, A.J. Regulation of glucose production by the liver. *Annual Review of Nutrition*, 19 (1999): 379-406.
doi:10.1146/annurev.nutr.19.1.379
- [7] Abdollahi, M., Saadat, M., Pournourmohammadi, S., Donyavi, M., Khorasani R, Amin, M. Alteration of rat hepatic glycogen phosphorylase and phosphoenolpyruvate carboxykinase activities by *Satureja khuzestanica* Jamzad essential oil. *Journal of Pharmacy Pharmaceutical Science*, 7(3) (2004b) 310-314.
- [8] Agius, L. New hepatic targets for glycemic control in diabetes mellitus. *Best Practise and Research, Clinical Endocrinology and Metabolism*, 21(4) (2007) 587-605
- [9] Zarmouh, K.S., Muftah, M., Viswanathan, S., and Kumar, P.G. Cause and effect of *Plumbago zeylanica* root extract on blood glucose and hepatic enzymes in experimental diabetic rats. *African Journal of Microbiology Research*, 4(24) (2010) 2674-2677.
- [10] Parka, H., Junga, U.J., Choa, S., Junga, H., Shimd, S., and Choi, M. Citrus unshiu peel extract ameliorates hyperglycemia and hepatic steatosis by altering inflammation and

- hepatic glucose-and lipid-regulating enzymes in db/db mice. *Journal of Nutritional Biochemistry*, 24(2) (2013) 419-427.
- [11] Farsi, E., Ahmad, M., Hor, S.Y., Ahamed, M.B.K., Yam, M.F., and Asmawi, Z.M. Standardized extract of *Ficus deltoidea* stimulates insulin secretion and blocks hepatic glucose production by regulating the expression of glucose-metabolic genes in streptozotocin-induced diabetic rats. *BMC Complementary and Alternative Medicine*, 14 (2014) 220.
- [12] Kolawale OT, Akanji MA. Effects of extract of leaves of *Newbouldia laevis* on the activities of some enzymes of hepatic glucose metabolism in diabetic rats. *African Journal of Biotechnology*, 13(22) (2014) 2273-228.
- [13] Hall, J.B., and Waljer, D.H. *Balanites aegyptiaca* Del. A Monograph School of Agricultural and Forest Science. Banger: University of Wales: (1991):1-2.
- [14] Hall, J.B. Ecology of a key African multipurpose tree species *Balanites aegyptiaca* Del. The state of knowledge. *Forest Ecological Management*, 50 (1992) 1-30.
- [15] Chothani D L and Vaghasiya HU. A review on *Balanites aegyptiaca* Del (desert date): phytochemical constituents, traditional uses, and pharmacological activity. *Pharmacogen Rev.* 5(9) (2011) 55-62.
- [16] Samir AM, Zaahkoug S, Rashid ZA, Mattar AF. Anti – diabetic properties of water and ethanolic extract of *Balanites aegyptiaca* fruits flesh in senile diabetic rats. *Egyptian Journal of Hospital Medicine*, 10 (2003): 90108.
- [17] George DH, Ali HK, El Abbas OA. Evaluation of the biological activity of *Balanites aegyptiaca* Del Saponin in the control of type 11 diabetes mellitus on rats and the growth of *Escherichia coli*. *Ahfad J. Women Change*, 23 (2006) 2.
- [18] Eman Helal GE, Abd El-Wahab SM, El Refaey H, Mohammad AA. Antidiabetic and antihyperlipidemic effect of *Balanites aegyptiaca* Seeds (Aqueous Extract) on diabetic rats. *The Egyptian Journal of Hospital Medicine*, 52 (2013) 725–739.
- [19] Ezzat SM, Abdel Motaal A, El-Awdan SAW. In vitro and in vivo antidiabetic potential of extract and a furostanol saponin from *Balanites aegyptiaca*. *Pharmaceutical Biology*, 55(1) (2017) 19311936.
- [20] Motaal AA, Shaker S, Haddad PS. Antidiabetic activity of standardized extract of *Balanites aegyptiaca* Fruits using cellbased bioassays. *Parmacognosy Journal*, 4(30) (2012) 20-24. 11.
- [21] Gad MZ, El-Sawalhi MM, Ismail MF, ElTanbouly ND. Biochemical study of the anti-diabetic action of the Egyptian plants Fenugreek and *Balanites*. *Molecular and Cellular Biochemistry*, 281 (2006) 173–183.
- [22] Gawade B, Farooqui M. Investigation of phytochemical and alpha amylase inhibition activity of *Balanites aegyptiaca* leaves. *Research Journal of Pharmaceutical, Biological and Chemical Sciences*. 9(1) (2018) 459-464.
- [23] Mhya, DH., Anigo KM., Umar IA., and Alegbejo, JO. Antihyperglycemic Effect of *Balanites aegyptiaca* Leaves Extract-Fractions in Streptozotocin-Induced Diabetic Rats. *Journal of Complementary and Alternative Medical Research*. 6(1) (2018) 1-12.

- [24] Pulok, K.M., Venkatesh, P., and Ponnusankar, S. Ethnopharmacology and integrative medicine – Let the history tell the future. *Journal of Ayurveda and Integrated Medicine*, 1(2) (2006) 100–109.
- [25] Oh, Y.S. Plant-Derived Compounds Targeting Pancreatic B- cells for the Treatment of Diabetes mellitus. *Evidence-Based Complementary and Alternative Medicine*, (2015): 1-12. doi.org/10.1155/2015/629863
- [26] Chawla, R., Thakur, P., Chowdhry, A., Jaiswal, S., Sharma, A., Goel, R. Evidence based herbal drug standardization approach in coping with challenges of holistic management of diabetes mellitus: a dreadful lifestyle disorder of 21st century. *Journal of Diabetes mellitus and Metabolic Disorders*, 12(1) (2013) 35-51.
- [27] Kazeem, M.I., Akanji, M.A., Yakubu, M.T., and Ashafa, A.O.T. Protective effect of free and bound polyphenol extract from Ginger (*Zingiber officinale* Roscoe) on the hepatic antioxidant and some carbohydrate metabolizing enzymes of streptozotocin-induced diabetic rats. *Evidence Based Complementary and Alternative Medicine*, 2013 (2013) 1-7.
- [28] Hosseini, A., Shafiee-Nick, R., and Ghorbani, A. Review: Pancreatic β - cell protection/regeneration with phytotherapy. *Brazilian Journal of Pharmaceutical Sciences*, 51(1) (2015) 1-16.
- [29] National Institute of Health (NIH). Principles of Laboratory Animal Care. NIH Publication. 1985; No. 85-23 Revised
- [30] Jung MY, Jeon BS, Bock JY. Free, esterified and insoluble bound phenolic acids in white and red ginsengs (*Panax ginseng* C.A. Meyer). *Food Chem.* 79 (2002) 105–111. 21.
- [31] Govorko D, Logendra S, Wang Y, Esposito D, Komarnytsky S, David R. Polyphenolic compounds from *Artemisia dracunculus* L. inhibit PEPCK gene expression and gluconeogenesis in an H4IIE hepatoma cell line. *Am J Physiol Endocrinol Metab.* 2007 293 1503–1510.
- [32] Gajdosik A, Gajdosikova A, Stefek M, Navarova J, Hozova R. Streptozotocin-induced experimental diabetes in male wistar rats. *Gen Physiol Biophys.* 18 (1999) 54-62.
- [33] Mhya DH, Amigo KM, Umar IA, Alegbejo JO. Evalaution of hypoglycemic potential of extracts of *Balanites aegyptiaca* parts. *Int. J of Innovative and Advanced Studies* 3(9) (2016) 135-138.
- [34] Goward, C.R., Hartwell, R., Atkinson, T., and Scawen, M.D. Enzymatic assay of glucokinase. Sigma Quality Control Test Procedure. (1986) 1-4
- [35] Hengartner H, Harris JI. Phosphofructokinase from *Bacillus stearothermophilus*. *Federation European Biochemical Society Letter*, 5(1975) 282.
- [36] Majumder AL, Eisenberg F Jr. In: Biswas T, Lahiri Majumder A, Guha G, Thakurt A, Mukherjee KL. Fructose-1, 6bisphosphatase in human fetal brain and liver during development. *J. Biosci.* 4(2) (1982) 167-173.

- [37] Chang HC, Lane MD. The enzymatic carboxylation of phosphoenolpyruvate. II. Purification and properties of liver mitochondrial phosphoenolpyruvate carboxykinase. *Journal of Biochemistry*. 241 (1966) 2413-2420.
- [38] Deutsch J. Glucose-6-phosphate dehydrogenase In: Bergmeyers Methods in Enzymatic Anal., 3rd Edition Beach, FL: Verlag Chemie. (1989):190.
- [39] Morgan HE, Parmeggiani A. Regulation of glycogenesis in muscle: Control of muscle glycogen phosphorylase activity. *Journal of Biological Chemistry*. 238(8) (1964) 2440-2445.
- [40] Baginski ES, Foa PP, Zak B. Glucose 6-phosphatase. In: Methods of enzymatic analysis. (Ed. Bergmeyer HU). New York: Verlag Chemie Weinheim. Acad. Press Inc. (1974) 737-764.
- [41] Danforth WH. Glycogen synthase assay. *Journal of Biological Chemistry*, (1965) 240: 588.
- [42] Pogson CI, Denton RM. Effect of alloxan diabetes, starvation and refeeding on glycolytic kinase activities in rat epididymal adipose tissue. *Nature*. 216 (1967) 156-157.
- [43] Duncan BD. Multiple range test for correlated and heteroscedastic means. *Biometrics*. 13 (1957) 359-364.
- [44] Mansour HA, Newairy AA. Amelioration of impaired renal function associated with diabetes by *Balanites aegyptiaca* fruits in streptozotocin-induced diabetic rats. *J Med Res Inst*. 21 (2000) 115-125.
- [45] Al-Malki AL, Barbour EK, Abullnaja KO, Moselhy SS. Management of hyperglycemia by ethyl acetate extract of *Balanites aegyptiaca* (Desert Date). *Molecules*. 20(8) (2015) 14425-14434.
- [46] Salwa AM, El Hadidi MN. Flavonoids of *Balanites aegyptiaca* (*Balanitaceae*) from Egypt. *Plant System Evolution*. 160(3) (1988) 153-158. 46.
- [47] Sarker SD, Bartholomew B, Nash RJ. Alkaloids from *Balanite aegyptiaca*. *Fitoterapia*. 71 (2000) 328-330.
- [48] Shafik NH, Shafek RZ, Michael HN, Eskander EF. Phytochemical study and antihyperglycemic effects of *Balanites aegyptiaca* kernel extract on alloxan induced diabetic male rats. *Journal of Chemistry and Pharmacy Research*, 8(3) (2016) 128-136.
- [49] Murphy ED, Anderson JW. Tissue glycolytic and gluconeogenic enzyme activities in mildly and moderately diabetic rats: Influence of *Tolbutamide* Administration. *Endocrinology*, 94(1) (1974) 27-34.
- [50] Latha M, Pari L. Antihyperglycemic effect of *Cassia auriculata* in experimental diabetes mellitus and its effects on key metabolic enzymes involved in carbohydrate metabolism. *Clinical Experiment in Pharmacology and Physiology*, 30(1-2) (2003) 38-43.
- [51] Nachar A, Vallerand D, Musallam S, Lavoie L, Badawi A, Arnason J. The action of antidiabetic plants of the Canadian James Bay Cree traditional pharmacopeia on key

- enzymes of hepatic glucose homeostasis. *Evidence-Based Complementary and Alternative Medicine*, (2013) 1-9.
- [52] Deepak KGK, Nageswara R, Neelapu R, Surekha C. Role of antidiabetic compounds on glucose metabolism – a special focus on medicinal plant: *Salacia* sps. *Medicinal Chemistry*, 4(3) (2014) 373-381.
- [53] Priyadarsini, R.L., Namatha, J.R., and Reddy, D.R.S. Glucokinase activators: a glucose sensor role in pancreatic islets and hepatocyte. *Pharmacy and Pharmaceutical Sciences*, 4(2) (2012) 81-87.
- [54] He L, Sabet A, Djedjos S, Miller R, Sun X, Hussain MA, Radovick S, Wondisford FE. Metformin and insulin suppress hepatic gluconeogenesis through phosphorylation of CREB binding protein. *Cell* 137(5) (2009) 635–646. 71
- [55] Meyer, F., Ipaktchi, M., and Clauser, H. Specific inhibition of gluconeogenesis by biguanides. *Diabetes mellitus*, 57(2) (2008) 306–314.
- [56] Viollet, B., Guigas, B., Garcia, N.S., Leclerc, J., Foretz, M., and Andreelli, F. Cellular and molecular mechanisms of metformin: an overview. *Clinical Science*, 122(6) (2012) 253-270.
- [57] Sharma, S.B., Rajpoot, R., Nasir, A., Prabhu, K.M., and Murthy, P.S. Ameliorative Effect of Active Principle Isolated from Seeds of *Eugenia jambolana* on Carbohydrate Metabolism in Experimental Diabetes mellitus. *Evidence-Based CAM* 78(9) (2011) 871-879.
- [58] Khanavi, M., Taheri, M., Rajabi, A., Fallah-Bonekohal, S., Baeri, M, Mohammadirad, A. Stimulation of Hepatic Glycogenolysis and Inhibition of Gluconeogenesis are the Mechanisms of Antidiabetic Effect of *Centaurea bruguierana* ssp. *belangerana*. *Asian Journal of Animal and Veterinary Advances*, 7 (2012) 1166-1174.
- [59] Khoramdelazad, H., Pourrashidi1, A., Hasanshahi, G., Vazirinejad, R., Hajizadeh, M., Mirzaei, M. Effect of the Iranian Walnut (*Juglans regia*) Leaves Extract on Gene Expression of Gluconeogenic and Glycogenolytic Enzymes in STZ-Induced Diabetic Rats. *International Journal of Current Research, Bioscience and Plant Biology*, 2(5) (2015) 47-55
- [60] Srivastava, S., Pant, K.K., and Lal, V.K.. Development, evaluation and quality control of new antidiabetic ayurvedic polyherbal combination. *African Journal of Pharmacy and Pharmacology*, (2014) 1146-1155.
- [61] Birudu RB, Naik MJ, Janardhan M. Ethanolic extract of *Passiflora foetida* and silver nanoparticles on carbohydrate metabolic enzymes of dextrose induced diabetic rats. *Journal of Biochemistry Biopharmacy and Biomedical Science*, 1(1) (2015) 12-19.
- [62] Jang SM, Kim MJ, Choi MS, Kwon EY, Lee MK. Inhibitory effects of ursolic acid on hepatic polyol pathway and glucose production in streptozotocin-induced diabetic mice. *Metabolic Clinical Experiment*. 59(16) (2010) 512–519.
- [63] Ramachandran V, Saravanan R. Efficacy of asiatic acid, a pentacyclic triterpene on attenuating the key enzymes activities of carbohydrate metabolism in streptozotocin-induced diabetic rats. *Phytomedicine*. 20(3-4) (2014) 230–236.

- [64] Gutierrez RMP. Evaluation of the hypoglycemic and hypolipidemic effects of triterpenoids from *Prosthechea michuacana* in STZ-induced type 2 diabetes mellitus in mice. *Pharmacologia* 4 (2013) 170–179. DOI: 10.5567/pharmacologia2013.170.179 70.
- [65] Xu, Y., Osborne, B.W., and Sanaon, R.C. Diabetes mellitus causes inhibition of glucose-6-phosphatase dehydrogenase via activation of PKA, which contribute to oxidative stress in rats' kidney cortex. *American Journal of Renal Physiology*, 289(5) (2005) 1040-1047.
- [66] Karuna, R., Bellamkonda, R., Singareddy, S.R., and Desireddy, S. Antihyperglycemic activity of *Catharanthus roseus* leaves powder in streptozotocin-induced diabetic rats. *Pharmacognosy Research* 2(3) (2010) 195-201.
- [67] Saralakumari, D., Ramesh, B., Karuna, R., Sreenivasa Reddy, S. and Sudhakara, G. Ethanolic extract of *Commiphora mukul* gum resin attenuates streptozotocin-induced alterations in carbohydrate and lipid metabolism in rats. *Excli Journal*, 12 (2013) 556-568.
- [68] Diaz-Flores, M., Ibanez-Hernandez, M.A., Galvan, R.E., Gutierrez, M., Duran-Reyes, G., Medina-Navarro, R. Glucose-6-phosphatase dehydrogenase activity and ADPH/NADP⁺ ratio in liver and pancreas are dependent on the severity of hyperglycemia in rat. *Life Science* 78(22) (2006) 2601-2607.
- [69] Jain, S.K. Glutathione and glucose-6-phosphatase dehydrogenase deficiency can increase protein glycosylation. *Free Radical and Biological Medicine*, 24(1) (1998) 197-201.
- [70] Atangwho, I.J., Khoo, B.Y., Umar, I.M., Ahmad, M., and Asmawi, M.Z. *Vernonia amygdalina* simultaneously suppresses gluconeogenesis and potentiates glucose oxidation via the pentose phosphate pathway in streptozotocin-induced diabetic rats. *BMC Complementary and Alternative Medicine*, 14 (2014) 426-439.